Identification of Glycoproteins from Mouse Skin Tumors and Plasma

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Abstract

Introduction Plasma has been the focus of testing different proteomic technologies for the identification of biomarkers due to its ready accessibility. However, it is not clear if direct proteomic analysis of plasma can be used to discover new marker proteins from tumors that are associated with tumor progression. In this paper, we reported that such proteins can be detected in plasma in a chemical-induced skin cancer model in mice.

Materials and Methods We analyzed glycoproteins from both benign papillomas and malignant carcinomas from mice using our recently developed platform, solid-phase extraction of glycopeptides and mass spectrometry, and identified 463 unique N-linked glycosites from 318 unique glycoproteins. These include most known extracellular proteins that have been reported to play roles in skin cancer development such as thrombospondin, cathepsins, epidermal growth factor receptor, cell adhesion molecules, cadherins, integrins, tuberin, fibulin, and TGF β receptor. We further investigated whether these tumor proteins could be detected in plasma from tumorbearing mice using isotope labeling and 2D liquid chromatography/matrix-assisted laser desorption/ionization tandem mass spectrometry.

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Department of Pathology, Johns Hopkins University, 1550 Orleans Street, CRBII, Room 3M-03, Baltimore, MD 21231, USA e-mail: hzhang32@jhmi.edu *Results and Discussion* Two tumor glycoproteins, Tenascin-C and Arylsulfatase B, were identified and quantified successfully in plasma from tumor bearing mice. This result indicates that analysis of tumor-associated proteins in tumors and plasma by a method using glycopeptide capture, isotopic labeling, and mass spectrometry can be used as a discovery tool to identify candidate tumor proteins that may be detected in plasma.

Keywords Cancer · Plasma · Tumor · Glycoprotein · Proteomics · Mass spectrometry · MALDI-TOF/TOF · MS/MS · Serum · Plasma · Biomarker

Introduction

Despite the great increase in understanding of cancer at the molecular level, cancer remains as the second most common cause of death in the USA. Survival rates for many common cancer types have changed little over the past two decades [1]. If cancer is detected early, prior to metastatic spread, survival rates are vastly improved [1]. For this reason, improvements in the ability to detect cancer early may significantly reduce mortality from cancer. Plasma has been the focus for applying different proteomic technologies for the identification of biomarkers for early detection due to its ready accessibility. These developments include depletion of the most abundant plasma proteins [2] and extensive fractionation of proteins or peptides prior to mass spectrometric analysis [3-5]. However, proteins discovered by serum profiling are often well-known, highabundance, classical serum proteins [6], not likely to be specifically derived from cancer tissue. Useful biomarkers for cancer detection in blood are those proteins released specifically from cancer tissues (overexpression of cancer proteins), indicators of a specific response of the host to cancer cells, or leaking of organ restricted proteins to blood due to structural changes in the microenvironment surrounding cancer cells (leaking of normal proteins such as PSA) [7]. Tumor proteins that are detectable in both benign and malignant tumors, as well as plasma can serve as candidate proteins for early detection of cancer. Detection of these proteins in plasma is critical to evaluate proteomic technologies for the biomarker discovery.

In an attempt to identify the proteins derived from cancerous tissue that are most likely to be present in blood, we employed our recently developed glycoproteomic analysis method using solid-phase extraction of N-linked glycopeptides (SPEG) [8-10]. The method has several advantages. First, most cell-surface and secreted proteins are glycosylated, and disease-associated glycoproteins (secreted by cells, shed from their surface, or otherwise released) are likely to enter the bloodstream and thus represent a rich source of potential disease markers [11]. Second, the reduction in complexity achieved by focusing on the glycoprotein subproteome in both tissues and plasma translates into favorable limits of detection, thus increasing the likelihood that the same polypeptide will be detectable in both tissue and serum [8, 12, 13]. Third, aberrant glycosylation is a fundamental characteristic of oncogenesis and tumor progression [14], and this method allows us to identify proteins changed in glycosylation but not necessarily changed in total protein abundance. Finally, specific mass-spectrometry-based methods and affinity reagents can be developed for the specific and sensitive detection of identified tissue proteins in plasma [15], selective isolation of specific proteins or peptides using affinity reagents [16], or the recently developed targeted approach using multiple reaction monitoring (MRM) [17-19].

The chemically induced two-stage mouse skin carcinogenesis model has been used for decades to study the genetic, molecular, and biologic basis of tumor development [20]. For example, the concepts of tumor initiation and promotion were derived from this model. In this model, the backs of 8-week-old mice treated with the carcinogen 7,12-dimethylben[a] anthracene (DMBA) followed by multiple treatments with the tumor promoter 12-o-tetradecanoylphorbol-13-acetate (TPA). Benign tumors (papillomas) develop after 8 weeks, and a small percentage of these progresses to malignant invasive carcinomas after a long latency [20]. The ability to quantify both benign and malignant tumor growth permits analysis of genes and environmental factors that affect tumor progression. More recently, the two-stage skin tumor model has been used to improve proteomic technologies for biomarker discovery using serum protein profiling [12]. We have identified several serum proteins for which the abundance is increased in correlation with the chemical induction of skin cancer in mice. However, these proteins are likely not markers for the specific diagnosis of skin cancer. A major advantage of this mouse skin carcinogenesis model is that plasma samples can be taken from mice before and after tumor development. As both benign and malignant tumors and plasma samples can be obtained from the same mice, this facilitates analysis of protein changes in plasma associated with tumor development.

In this paper, we report a two-step strategy for detection of tumor-associated proteins in plasma: the first step was to analyze extracellular proteins from normal skin, papillomas, and carcinomas and identify tumor-associated proteins; the second step was to detect the tumor-associated proteins in plasma using a tissue-targeted approach and isotope labeling [7]. Using our recently developed method of SPEG and mass spectrometry [8-10], we analyzed matched benign and cancerous tumors from four tumor-bearing mice as well as normal skin tissues from four control mice, and identified 463 unique N-linked glycosites from 318 glycoproteins. More than 40 identified glycoproteins were elevated in carcinomas. Two of the tumor-associated proteins, Tenascin-C and Arylsulfatase B, were further detected and quantified in plasma from the same cancerbearing mice using isotope labeling and 2D liquid chromatography/matrix-assisted laser desorption/ionization tandem mass spectrometry (LC-MALDI-MS/MS). This result indicates that direct proteomic analysis of tumors and plasma using glycopeptide capture, isotopic labeling, and mass spectrometry can be used to discover new cancer-derived proteins in plasma.

Method and Materials

Materials

Hydrazide resin and sodium periodate were from Bio-Rad (Hercules, CA, USA); PNGase F was from New England Biolabs (Ipswich, MA, USA); sequencing grade trypsin was purchased from Promega (Madison, WI, USA); C18 columns were from Waters (Milford, MA, USA); α -cyano-4-hydroxycinnamic acid (CHCA) was from Agilent (Palo Alto, CA, USA); iTRAQ reagent and mass calibration standards were purchased from Applied Biosystems (Foster City, CA, USA). All other chemicals were purchased from Sigma-Aldrich (St. Louis, MO, USA).

Tissues and Plasma from Chemical-Induced Mouse Skin Tumors

Skin tumors were induced in four NIH01a mice using the DMBA/TPA two-step protocol. A single dose of DMBA (Sigma; 25 mg in 200 ml of acetone) was applied to the shaved

backs of four 8-week-old mice. Initiated treated skin was promoted with TPA twice a week for 15 weeks. This gave rise to papillomas that were hyperplastic, well-differentiated, benign lesions consisting of keratinocytes together with stroma tissue. Papillomas appeared as early as 8 weeks after the first treatment of DMBA and continued to grow for the next several months. A small percentage of these benign papillomas (~20%) progressed to squamous cell carcinomas. All the mice were killed when carcinomas appeared in all four treated mice. Four littermate mice were left untreated for normal skin tissues. Papillomas and carcinomas, as well as normal skin from untreated mice were snap-frozen in liquid nitrogen. Retro-orbital bleeds were collected from each treated mouse before chemical treatment and after development of chemical-induced carcinomas. The only difference between the normal and cancer tissues is the chemical-induced cancer. Whole blood (0.25 ml) was collected from the retro-orbital sinus into a long (9 in.) sterile glass Pasteur pipet. The whole blood was placed in a K3EDTA-coated 1.5 ml microcentrifuge tube and centrifuged at 4°C for 5 min at 3,000 rpm. Plasma was collected, carefully avoiding cellular contamination. All tumor tissues and plasma were placed in cryovials and frozen in liquid nitrogen.

Peptide Extraction from Skin Tumor Tissues

Frozen tumor tissues (100 mg each) were sliced into 1- to 3-mm³ thickness and incubated in 200 µl of 5 mM phosphate buffer and vortexed for 2-3 min. Then, the samples were sonicated for 5 min in an ice-water bath. Trifluoroethanol (TFE, 200µl) was added to the sample and incubated at 60°C for 2 h followed by sonication for 2 min. Protein disulfide bonds were reduced by 5 mM tributylphosphine with 30-min incubation at 60°C. Iodoacetamide (10 mM) was applied to the mixture and incubated in the dark at room temperature for another 30 min. The samples were diluted fivefold with 50 mM NH₄HCO₃ (pH 7.8) to reduce the TFE concentration to 10% prior to the addition of trypsin at a ratio of 1:50 (w/w, enzyme/protein). Samples were digested at 37°C overnight with gentle shaking. The precipitate was discarded by centrifuge. Silver staining was used to test the effect of tryptic digestion. Four milligrams of total peptides from each sample was extracted from each tissue. Two milligrams of total peptide was used to extract N-linked glycopeptides, according to the following steps.

Peptide Extraction from Plasma

Plasma (20 μ l) was added to 90 μ l 8 M urea in 0.4 M NH₄HCO₃, 0.1% (*w*/*v*) sodium dodecyl sulfate (SDS) solution (pH 8.3) and 10 μ l 120 mM tris(2-carboxyethyl) phosphine in dH₂O freshly prepared and incubated at 60°C for 1 h. Proteins were alkylated by adding 10 μ l 160 mM

iodoacetamide and incubated at room temperature in the dark with shaking for another 30 min. Samples were diluted by trypsin digestion buffer (100 mM NH₄HCO₃, pH 8.3) to make the concentration of urea less than 2 M. Forty microliters of trypsin ($0.5\mu g/\mu l$) was added to digest protein at 37°C overnight. SDS-polyacrylamide gel electrophoresis (SDS-PAGE) and silver staining were employed to check whether trypsin digestion was complete.

Glycopeptide Capture from Tissue or Plasma

N-glycopeptides were isolated from peptides using SPEG [9]. The enriched N-linked glycopeptides were concentrated by C18 columns and dried down and resuspended in 40 μ l 0.4% acetic acid prior to MS analysis.

Isotope Labeling of Peptides

The amount of glycopeptide was determined by bicinchoninic acid assay (Bio-Rad) prior to isotope labeling. Glycopeptides (1 µg) from plasma of the retro-orbital bleeds before and after chemical-induced cancer and tumor tissues were dried and resuspended in 20 µl of 50% dimethylformamide, 40% H₂O, 10% pyridine. Five microliters 10 mg/ml d0¹³C0, d4¹³C0, and d4¹³C4 succinic anhydride solution was added to glycopeptide samples and reacted at room temperature for 1–2 h, then followed by C18 clean up to remove access succinic anhydride [8].

Mass Spectrometry Analysis

The peptides and proteins were identified using MS/MS analysis using an LTQ ion trap mass spectrometer (Thermo Finnigan, San Jose, CA, USA). Glycopeptides (1 µg) were injected into a peptide cartridge packed with C18 resin and then passed through a 10 cm×75 µm i.d. microcapillary high-performance liquid chromatography (µLC) column packed with C18 resin. The effluent from the µLC column entered an electrospray ionization source in which peptides were ionized and passed directly into the mass spectrometer. A linear gradient of acetonitrile from 5% to 32% over 100 min at flow rate of ~300 nl/min was applied. During the LC-MS mode, data were acquired between *m/z* of 400 and 2,000. Each sample was analyzed three times to increase the number of spectra used for spectral count.

Succinic-anhydride-labeled peptide (5 μ g) was analyzed by 2D Nano LC (Eksigent, Dublin, CA, USA) and MALDI-tandem time of flight (MALDI-TOF/TOF; Applied Biosystems). Briefly, online integration of 15-cm-long 300 μ m strong cation exchange column (SCX) with 15-cm-long 300 μ m of C18-reverse phase liquid chromatograph (RPLC) was employed. Four SCX fractions of 0, 5, 50, and 500 mM KCl and 3–45% linear acetonitrile gradient (containing 0.1% TFA and acetonitrile) of RPLC for each fraction were applied before analysis by MALDI-TOF/TOF. Peptides eluted from columns were directly mixed with α -Cyano-4-hydroxycinnamic acid and spotted on a MALDI target plate with 768 spots followed by analysis by MS and MS/MS using ABI4800 MALDI-TOF/TOF.

Data Analyses

Peptide identifications-MS/MS spectra from LTQ were searched with SEQUEST [21] against a mouse protein database (the International Protein Index mouse protein database, version 3.13). The precursor mass tolerance is set as 3.0 Da. Other parameters of database searching are modified as follows: oxidized methionines (add Met with 16 Da), a (PNGase F-catalyzed) conversion of Asn to Asp (add Asn with 1 Da), and Cys modification (add cysteine with 57 Da). The output files were evaluated by INTERACT and PeptideProphet [22, 23]. The criterion of PeptideProphet analysis is the probability score ≥ 0.9 so that low probability protein identifications can be filtered out.

Identifying tissue-derived peptides in plasma from MALDI-TOF/TOF (ABI 4800) was performed using GPS Explorer software (version 3.6). MS/MS spectra were searched against NCBInr database. GPS searches were carried out at a 0.2-Da precursor mass tolerance, a 0.6-Da fragment mass tolerance, trypsin as digestion enzyme. In addition to the modifications for Met, Asp, and Cys that were used in LTQ MS/MS spectra analyses as described above, N termini of peptides and Lys are modified by succinic anhydride (100 Da for d0¹³C0, 104 Da for d4¹³C0, and 108 Da for d4¹³C4).

Results and Discussion

Strategy of the Method

The objective of this study was to use N-linked glycopeptide isolation, isotopic labeling, and LC-MS to identify skin-cancer-related extracellular proteins and determine if these proteins could be detected in plasma from tumorbearing mice. This strategy is based on the fact that most extracellular proteins are glycoproteins, and extracellular proteins from cancer are most likely to be detected in plasma due to the fact that they are likely to be secreted by cells or shed from the cell surface to enter the blood stream.

The strategy is schematically illustrated in Fig. 1 and consists of four steps: (1) peptide extraction from tissue or plasma; (2) glycopeptide extraction: peptides that contain N-linked carbohydrates in extracellular proteins were isolated in their de-glycosylated form using a recently described solid-phase capture-and-release method [9, 10]; (3) identification and quantification analysis of glycopeptides isolated from normal skin, papillomas, and carcinomas: isolated peptides were analyzed by LC-MS/MS, and the peptides were identified and quantified using a database search [21] and spectral count; (4) detection of tissuederived proteins in plasma. Glycopeptides from plasma samples taken from mice before and after development of skin tumors and tumor tissues were labeled with $d0^{13}C0$, d4¹³C0, and d4¹³C4 succinic anhydride, respectively. The peptides containing d4¹³C0 and d4¹³C4 pairs indicated the tumor-derived peptides detected in plasma from tumorbearing mice, and they were selected for MS/MS analysis for peptide identifications.

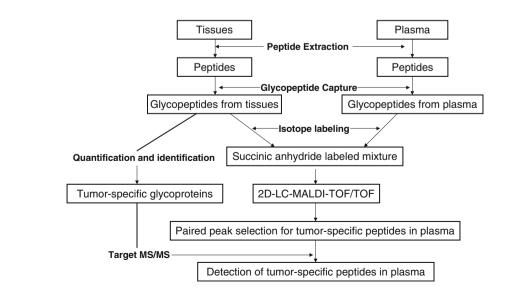


Fig. 1 Flow chart for detection of tumor-specific proteins in plasma

Identification of Proteins from a Mouse Model of Skin Cancer

To detect tumor-specific proteins in plasma, we first identified tumor-associated proteins from carcinomas and papillomas. These tumor-associated proteins are likely to be secreted or shed to the blood stream and fall into the detection range of current proteomic methodology.

To identify extracellular proteins from mouse skin tumors, four tissue samples each from normal skin, benign papillomas, and malignant carcinomas were collected to generate pooled normal, benign, and cancer tissues. Proteins were extracted from homogenized frozen tissues and digested to peptides. Glycopeptides were then captured using SPEG from each tissue. The N-linked glycopeptides were analyzed by LC-MS/MS by three repeated analyses for each sample. The MS/MS spectra were used to search protein databases using SEQUEST [21]. There were a total of 4,764 peptide identifications with PeptideProphet of at least 0.9 (with error rate of 0.007) from all the tissues. Ninety percent of these identifications (4,284 identifications) contained a consensus N-linked glycosylation motif (N-X-S/ T, X is any amino acid except proline). These identifications were from 463 unique glycosylation sites representing 318 unique glycoproteins (Table 1). This indicated that the procedure was specific to N-linked glycoproteins. Therefore, we limited our subsequent analysis solely to the identified peptide sequences that contained at least one such consensus motif in order to reduce false positive rates. Since tissues are vascularized and some proteins identified from tissues are from contamination by common circulating blood proteins [13, 24], we next examined the glycoproteins identified from tissues to determine glycoproteins identified from tissues that were also identified from the normal mouse plasma [10, 25], and 59 glycoproteins were previously identified from normal mouse plasma and were not included for further study of skin cancer tissues.

To identify skin tumor-specific proteins, we compared the glycoproteins identified from normal skin, benign, and malignant tumors. Despite the same amount of glycopeptides from each tissue analyzed with the same procedures, the number of unique glycosites identified from different tissues was different. A total of 405 glycosites were identified in cancer tissue, while 252 in benign tissue and 112 in normal skin, when using PeptideProphet score of ≥ 0.9 . The number of glycoproteins identified from papillomas and carcinoma was higher than that of normal tissue. This could be caused by the increased expression of glycoproteins in tumor tissues. A similar observation was also reported from the proteomic analysis of tryptic peptides in a mouse breast cancer model [24].

To determine the glycoprotein changes associated with cancer development, we calculated the relative protein abundance using the number of redundant MS/MS spectra from the same glycoprotein in different tissues [26]. To eliminate the spectral count due to random events, only proteins identified with at least three spectra were included for quantitation. A number of proteins identified in this study were only detected in tumor tissues (benign or malignant) but not in normal tissues (the ratio of such proteins was arbitrary assigned to 100, Table 2). Among the 111 proteins identified with spectral count ratio of at least three folds in cancer or benign tumor tissues comparing to normal tissues, 47 proteins (Table 2) were increased at least three folds in cancer tissues compared to benign tissues. Some of these have been reported to play roles in skin cancer development. These include known extracellular proteins such as thrombospondin, cathepsins, epidermal growth factor receptor, cell adhesion molecules, cadherins, integrins, tuberin, fibulin, and TGFB receptor. Tenascin-C is an extracellular matrix glycoprotein and plays multiple functions in cell adhesion, migration, growth, and angiogenesis [27, 28]. Tenascin-C has many cell surface receptors, such as integrin and epidermal growth factor receptor, which may affect genome stability associated with interference with genome safeguard functions and escape from cell cycle checkpoints [28]. Tenascin-C has 20 potential N-linked glycosylation sites, but only one glycosylation site (LLQTAEHN#ISGAER, Table 1) has been identified previously (Swiss-Prot Protein knowledgebase, http://us.expasy. org/sprot). In this study, eight N-linked glycosites including the previously identified site were identified in carcinomas (Table 1). They showed increased expression in carcinomas compared to papillomas (Table 2). This observation indicated that Tenascin-C might have increased its glycosylation or abundance during tumor development. In addition, 20 glycoproteins were identified in skin cancer only (Table 2), and these proteins might be used as protein markers to discriminate between the malignant and benign tumors. An example is Arylsulfatase B. In this study, Arylsulfatase B was identified three times only in malignant tissues with two unique glycosylation sites. Arylsulfatase B is a lysosomal enzyme and can degrade proteoglycans in the extracellular matrix and basement membrane. In this way, proteoglycans can obstruct the spread of cancer cells. Therefore, Arylsulfatase B could play a key role in accelerating cancer cell migration [29].

Here, we determined the relative abundance of glycosylated proteins using identified glycosylated peptides. However, glycosylation for individual glycosites from the same protein might be different and can be determined by quantitative analysis of each glycosite. In addition, changes in glycan structure that may be important to the disease cannot be determined by this method, and specific enrichment of glycopeptides with certain glycan structure is needed.

Table 1 Identified N-linked glycoproteins and glycosites

IPI	Protein name	Р	Identified sequences
IPI00120245	Integrin alpha-V	1	K.AN#TTQPGIVEGGQVLK.C
IPI00120245	Integrin alpha-V	1	R.TAADATGLQPILNQFTPAN#VSR.Q
PI00127447	Lysosome membrane protein II	1	R.N#QSVGDPNVDLIR.T
PI00127447	Lysosome membrane protein II	1	T.GEDNYLN#FSK.I
PI00127447	Lysosome membrane protein II	1	R.TMVFPVMYLN#ESVLIDK.E
PI00127447	Lysosome membrane protein II	1	R.YKVPAEILAN#TSENAGF.C
PI00322447	RA175	1	K.VSLTN#VSISDEGR.Y
PI00322447	RA175	1	R.FQLLN#FSSSELK.V
PI00118413	Thrombospondin 1	1	L.DNNVVN#GSSPAIR.T
PI00118413	Thrombospondin 1	1	K.VSCPIMPCSN#ATVPDGECCPR.C
PI00118413	Thrombospondin 1	1	W.PNENLVCVAN#ATYHCK.K
	Cadherin-22	0.98	R.ETAGWHN#ITVLAMEADN.H
PI00154057	Protocadherin 1	0.99	N.DNAPFITAPSN#TSHR.L
PI00126090	Integrin alpha-3	1	I.AMN#YSLPLR.M
PI00126090	Integrin alpha-3	1	W.LECPLPDTSN#ITN#VTVK.A
PI00132474	Integrin beta-1	1	R.NPCTSEQN#CTSPFSYK.N
PI00132474	Integrin beta-1	1	R.KEN#SSEICSNNGECVCGQCVCR.K
PI00132474	Integrin beta-1	1	K.DTCAQECSHFN#LTK.V
PI00227969	Integrin alpha-6	1	K.YQTLN#CSVNVR.C
PI00227969	Integrin alpha-6	0.91	R.VEQKN#NTFFDMNIF.E
PI00320605	Integrin beta-2	1	K.LN#FTGPGEPDSLR.C
PI00320605	Integrin beta-2	0.99	Y.LRPGQAAAFN#VTFR.R
PI00415773	Integrin alpha-M	1	R.TPVLN#CSVAVCK.R
	Integrin alpha-M	1	V.GGPQDFN#MSVTLR.N
	Integrin alpha-M	1	R.LN#YTLVGEPLR.S
PI00132067		1	Y.QLPGCHGN#FSDAEEGDSER.Q
PI00132067	Fibulin-2	1	K.DLDECALGTHN#CSEAETCHNIQGSFR.C
PI00132067	Fibulin-2	1	K.SCVAGVMGAKEGETCGAEDN#DTCGVSLYK.A
PI00223769	CD44 antigen	1	R.TEAADLCQAFN#STLPTMDQMK.L
	Prostate stem cell antigen	1	R.DCLNVQN#CSLDQHSCFTSR.I
	Translocon-associated protein alpha, muscle specific isoform	1	K.DLNGNVFQDAVFN#QTVT.V
PI00110852	Translocon-associated protein alpha, muscle specific isoform	1	R.YPQDYQFYIQN#FTALPLNTVVPPQR.Q
PI00112326	Epithelial membrane protein 1	1	K.N#CTGGNCDGSLSYGNEDAIK.A
	Myeloperoxidase	1	R.ALMPFDSLHDDPCLLTN#R.S
	Cathepsin D	1	K.YYHGELSYLN#VTR.K
	Cathepsin D	1	K.N#GTSFDIHYGSGSL.S
	Cathepsin L	1	R.AEFAVAN#DTGFVDIPQQEK.A
	Tenascin-C	1	L.EADTTQTVQN#LTVPGGLR.S
	Tenascin-C	1	R.EPEIGNLN#VSDVTPK.S
	Tenascin-C	1	R.LLQTAEHN#ISGAER.T
	Tenascin-C	1	N.NVEAAQN#LTVPGSLR.A
	Tenascin-C		N.NVETAHN#FTVPGNLR.A
	Tenascin-C	1	R.ESGLN#MTLPEENQPVVFNHIYNIK.L
	Tenascin-C	1	K.ASTEEVPSLEN#LTVT.E
	Tenascin-C	1	R.LN#YSLPTGQSMEVQLPK.D
	Carcinoembryonic antigen-related cell adhesion molecule 1	1	R.FVPNSNMN#FTGQAYSGR.E
PI00108535	Carcinoembryonic antigen-related cell adhesion molecule 1	1	K.N#ITVLEPVTQPFLQVTN#TTVK.E
0100212/20	CEA-related cell adhesion molecule 2	1	R.TLTLLN#VTR.N
	Plasma protease C1 inhibitor	1	R. TLTLLN#VTR.N R.DTYVN#ASQSLYGSSPR.V
	Plasma protease C1 inhibitor	1	
	Collagen alpha 1(V) chain	1	K.VGQLQLSHN#LSFVIVVPVFPK.H K.VYCN#FTAGGSTCVFPDKK.S
	GPI-anchored metastasis-associated protein homolog	1	A.N#VTVSLPVR.G
	VIT 1-AUCHOICU IUCIASIASIS-ASSOCIATED DIOTEIII DOMOTO	1	A.IN# VIV SLEVE.U

IPI

IPI00130249 IPI00130486 IPI00130486 IPI00132600 IPI00132600 IPI00131881 IPI00130342 IPI00130342 IPI00133082 IPI00134549 IPI00134549 IPI00134549 IPI00121430 IPI00122272 IPI00122272 IPI00122493 IPI00122493 IPI00123342 IPI00123342 IPI00123342 IPI00123831 IPI00123831 IPI00123831 IPI00224728 IPI00462199 IPI00462199 IPI00462199 IPI00308609

IPI00308990 IPI00308785 IPI00308971 IPI00308971 IPI00124836 IPI00124836 IPI00122737 IPI00122737 IPI00119063 IPI00119063 IPI00119063 IPI00119063 IPI00119063 IPI00119063 IPI00124265 IPI00124265 IPI00129304 IPI00129304 IPI00153959 IPI00153959 IPI00316575 IPI00126769 IPI00121190 IPI00320420 IPI00320420 IPI00406459 IPI00406459

Protein name	Р	Identified sequences
GPI-anchored metastasis-associated protein homolog	1	K.CQGSMPPVVNCYN#ASGR.V
FK506-binding protein 9	1	R.YHYN#GTLLDGTLFDSSYSR.N
FK506-binding protein 9	1	R.YHYN#GTFLDGTLFDSSHNR.M
Niemann-Pick C1 protein	1	R.LYN#VTHQFCN#ASVMDPTCVR.C
Niemann-Pick C1 protein	1	R.LIASN#ITETMR.S
ADAM 10	1	R.IN#TTSDEKDPTNPFR.F
Lymphocyte antigen 6 complex locus G6C protein	1	K.LGLNYN#TTCCDK.D
Lymphocyte antigen 6 complex locus G6C protein	1	R.EVFN#ETNHK.L
CD177 antigen	1	K.VQGCMAQPDCNLLN#GTQAI.G
Lysosome-associated membrane glycoprotein 2	1	A.LIVN#LTDSK.G
Lysosome-associated membrane glycoprotein 2	1	K.VPFIFNINPATTN#FTGSCQPQSAQLR.L
Lysosome-associated membrane glycoprotein 2	1	K.EVNVYMYLAN#GSAFN#ISNK.N
Collagen alpha 1(XII) chain	1	K.EAGN#ITTDGYEILGK.L
Extracellular matrix protein 1	1	K.QIPGLIQN#MTVR.C
Extracellular matrix protein 1	1	R.NVALVAGDTGN#ATGLGEQGPTR.G
FK506-binding protein 10	1	R.YHYN#CSLLDGTR.L
FK506-binding protein 10	1	R.YHYN#GSLMDGTLFDSSYSR.N
Hypoxia up-regulated 1	1	R.VFGSQN#LTTVK.L
Hypoxia up-regulated 1	1	R.LSALDNLLN#HSSIFLK.G
Hypoxia up-regulated 1	1	K.EN#GTDAVQEEEESPAEGSK.D
SDR1 protein	1	K.ENGVFEEISN#SSGR.F
SDR1 protein	1	R.FFITNKEN#YTEL.S
SDR1 protein	1	R.ESLLPVTLQCN#LTSSSH.T
Cd63 antigen	1	K.DRVPDSCCIN#ITVGCGNDFK.E
Basigin	1	K.TSDTGEEEAITN#STEANGK.Y
Basigin	1	K.TQLTCSLN#SSGVDIVGHR.W
Basigin	1	K.SQLTISNLDVNVDPGTYVCN#ATNAQGTTR.E
VESICULAR INTEGRAL-MEMBRANE PROTEIN	1	R.VFPYISVMVNN#GSLSYDHSK.D
VIP36		
Monocyte differentiation antigen CD14	1	R.N#PSPDELPQVGN#LSLK.G
Prostaglandin G/H synthase 2	1	R.TGFYGEN#CTTPEFLTR.I
Cation-independent mannose-6-phosphate receptor	1	K.ISTN#ITLVCKPGDLESAPVLR.A
Cation-independent mannose-6-phosphate receptor	1	R.SLLEFN#TTMGCQPSDSQHR.I
Beta-sarcoglycan	1	R.ITSN#ATSDLNIK.V
Beta-sarcoglycan	0.99	I.ILN#GTVMVSPTR.L
222 kDa protein	1	R.QAEEAEEQANTN#LSK.F
222 kDa protein	0.98	R.VQLLHSQN#TSLINQKK.K
AM2 receptor	1	K.LTSCATN#ASMCGDEAR.C
AM2 receptor	1	K.LNLDGSN#YTLLK.Q
AM2 receptor	1	A.VAN#DTNSCELSPCR.I
AM2 receptor	1	R.MGCQHHCVPTPSGPTCYCN#SSFQLE.A
AM2 receptor	0.99	R.GVTHLN#ISGLK.M
AM2 receptor	1	R.FN#STEYQVVTR.V
Latent transforming growth factor beta binding protein 4	1	R.N#ATSVDSGAPGGAAPGGPGFR.A
Latent transforming growth factor beta binding protein 4	1	R.CTPACDPGYQPTPGGGCQDVDECRN#R.S
Collectin sub-family member 12	1	R.HTDDLTSLN#NTLVNIR.L
Collectin sub-family member 13	1	K.ETLQN#NSFLITTVN#K.T
Stabilin-1	1	H.ADLISN#MSQDELAR.I
Stabilin-1	1	K.GFVDN#MTLSGPDLELH.A
Cathepsin K	1	Y.VGQDESCMYN#ATAK.A
Cathepsin F		K.VYIN#DSVELSR.N
Epidermal growth factor receptor	1	R.DIVQNVFMSN#MSMDLQSHPSSCPK.C
Clusterin	1	R.QELN#DSLQVAER.L
Clusterin		K.MLN#TSSLLEQLNDQFNWVSQLAN#LTQGEDK.Y
Arylsulfatase B	1	H.EACAPIESLN#GTR.C
Arylsulfatase B	1	R.IYAGMVSLMDEAVGN#VTK.A

Ы	Protein name	Р	Identified sequences			
PI00409393	Latent transforming growth factor beta binding protein, isoform 1L	1	R.YGQEQGTAPFQVSN#HTGR.I			
PI00409393	Latent transforming growth factor beta binding protein, isoform 1L	1	Y.NLNDASLCDNVLAPN#VTK.Q			
PI00409393	Latent transforming growth factor beta binding protein, isoform 1L	0.91	K.VCTN#GSCTNLEGSYM.C			
PI00108535	Carcinoembryonic antigen-related cell adhesion molecule 1	1	R.EIIYSN#GSLLFQMITMK.D			
PI00117424	Intercellular adhesion molecule 2	1	K.IN#CSTNCAAPDMGGLETPTNK.I			
	N-CAM 180 of Neural cell adhesion molecule 1, 180 kDa isoform	1	R.DGQLLPSSN#YSNIK.I			
PI00406901	Platelet/endothelial cell adhesion molecule	1	K.EETVLSQYQN#FSK.I			
	Integrin alpha-5	1	K.VTGLSN#CTSN#YTPN.S			
	Integrin beta 4 Isoform 2	1	K.TCN#CSTGSLSDTQPCLR.E			
	Integrin alpha 1	1	K.DSCESNQN#ITCR.V			
PI00230432			H.SYN#SSLETIFIK.R			
	OX-2 membrane glycoprotein	1	K.GTGTGIEN#STESHFHSN#GTTSVTSILR.V			
	PTK7 protein tyrosine kinase 7		R.MHIFQN#GSLVIH.D			
	TGF-beta receptor type III	1	R.AGVVVFN#CSLR.Q			
	Cell surface glycoprotein OX2 receptor	1	W.SPDGDCVTTSESHSN#GTVTVR.S			
	Transmembrane 9 superfamily protein member 3	1	R.IVDVN#LTSEGK.V			
	Thrombospondin-3	1	R.LGFLGN#QSQGCVPAR.T			
	Mama protein	1	R.ALGYEN#ATQALGR.A			
	-					
	Mama protein	1	K.GLN#LTEDTYKPR.L			
	Lysosomal membrane glycoprotein 1	1	R.LN#MTLPDALVPTFSISN#HSLK.A			
	Lysosomal membrane glycoprotein 2	1	K.N#VTVVLR.D			
	Similar to KALLIKREIN 9	1	R.LTPAVQPLN#LTESRPPVGTQ.C			
	Glandular kallikrein KLK13	1	K.ILN#GTN#GTSGFLPGGYTCLPH.S			
PI00116993			A.PKQGLN#NSPPVK.E			
	Mucin and cadherin-like protein	1	R.VTN#SSEFMMNK.D			
	Stromal interaction molecule 1	1	R.LAVTN#TTMTGTVLK.M			
	Methylated-DNA- protein-cysteine methyltransferase containing protein		M.ETTSLLLCIGN#NSSGIRSRHR.S			
	Glucosylceramidase	1	R.DLGPALAN#SSHDVK.L			
	Enabled protein homolog	1	W.ERTNTMN#GSK.S			
	Laminin, beta 2	1	L.ASGN#VSGGVCDGCQHNTAGR.H			
	Thy-1 membrane glycoprotein	1	K.VLTLAN#FTTK.D			
	Ig gamma-2A chain C region, membrane-bound form	1	R.EDYN#STLR.V			
	Elongation of very long chain fatty acids protein 4	1	T.AFN#DTVEFYR.W			
	Similar to METASTASIS-ASSOCIATED GPI- ANCHORED PROTEIN	1	R.MNIGN#FSVPVYIR.T			
	Lysosomal alpha-glucosidase	1	R.GVFITN#ETGQPLIGK.V			
	Copper homeostasis protein cutC homolog		R.N#SSVAMGASLAHSEYSLK.V			
	Plasma kallikrein	1	K.LQTPLN#YTEFQKPICLPSK.A			
PI00113797			W.FN#LTGQDYVIK.I			
	Basement membrane-specific heparan sulfate proteoglycan core protein	0.98	K.LTVPSSQN#SSFR.L			
	Basement membrane-specific heparan sulfate proteoglycan core protein	1	R.SLTQGSLIVGNLAPVN#GTSQGK.F			
	Basement membrane-specific heparan sulfate proteoglycan core protein	1	R.VAQQDSGQYICN#ATNSAGH.T			
	Desmocollin-3		K.AN#FTILK.G			
	Eosinophil peroxidase	0.99	F.DNLHEDPCLLTN#R.S			
	Complement factor B	1	K.IVLDPSGSMNIYLVLDGSDSIGSSN#FTGAK.R			
'I00114065	Complement factor B	0.94	R.SPFYN#LSDQI.S			
	Prothrombin		R.WVLTAAHCILYPPWDKN#FTENDLLVR.I			

IPI	Protein name	Р	Identified sequences
IPI00114206	Prothrombin	1	R.ITDNMFCAGFKVN#DTK.R
IPI00400016	Laminin gamma-1 chain	1	K.LLNN#LTSIK.I
IPI00400016	Laminin gamma-1 chain	1	R.TLAGEN#QTALEIEELNR.K
IPI00400016	Laminin gamma-1 chain	1	L.SYGQN#LSFSFR.V
IPI00400016	Laminin gamma-1 chain	1	R.KYEQAKN#ISQDLEKQ.A
IPI00317340	Lactotransferrin	1	I.PMGLLAN#QTR.S
IPI00317340	Lactotransferrin	1	K.N#SSNFHLNQLQGLR.S
IPI00113539	Fibronectin	1	R.DQCIVDDITYNVN#DTFHK.R
IPI00113539	Fibronectin	1	K.LDAPTNLQFVN#ETDR.T
IPI00113539	Fibronectin	1	R.HEEGHMLN#CTCFGQGR.G
IPI00119818	Inter alpha-trypsin inhibitor, heavy chain 4	1	K.AFITN#FSMIIDGVTYPGVVK.E
IPI00119818	Inter alpha-trypsin inhibitor, heavy chain 5	1	R.GLMLLLN#DTQHFSNNVK.G
IPI00114256	Synaptophysin-like protein	1	K.N#QTVTATFGYPFR.L
IPI00114319	Extracellular superoxide dismutase [Cu-Zn]	1	R.LEAYFSLEGFPAEQN#ASNR.A
IPI00114641	CD98 heavy chain	1	K.LMNAPLYLAEWQN#ITK.N
	Suppressor of tumorigenicity 14	0.99	R.VIN#QTTCEDLMPQQITPR.M
	HMW of Kininogen-1	1	K.HSIEHFNN#NTDHSHLFTLR.K
	HMW of Kininogen-1	1	T.YTIVQTN#CSK.E
	HMW of Kininogen-1	1	K.IAN#FSQSCTLYSGDDLVEALPKPCPGCPR.D
	Ectonucleoside triphosphate diphosphohydrolase 2	1	R.LLN#LTSPEATAK.V
IPI00115516		1	R.FN#STLGPSEEQEK.N
IPI00115530	Beta-hexosaminidase beta chain	1	K.TQVFGPVDPTVN#TTYA.F
IPI00115762	Neural cell adhesion molecule L1	1	K.EQLFFN#LSDPELR.T
IPI00115817	PREDICTED: similar to ribosomal protein L21	0.95	K.TGRVYN#VTOHAMGIIVNK.O
	TROP2 protein	1	R.AFN#HSDLDSELR.R
	Corticosteroid-binding globulin	1	K.DLFTN#QSDFADTTK.D
	Corticosteroid-binding globulin	1	R.EEDFYVN#ETSTVK.V
	Corticosteroid-binding globulin	1	K.VPMMVQSGN#ISYFR.D
	Corticosteroid-binding globulin	1	R.GSTQYLENLGFN#MSK.M
	p130Cas-associated protein		R.RQVDEGMWPPPNNLLN#QSPK.K
	Laminin alpha-5 chain	1	R.QLLAN#SSALEETILGHQGR.L
	Laminin alpha-5 chain	1	H.N#FSGCISNVFVQR.L
	Complement factor D	1	K.LSQN#ASLGPHVRPLPLQYEDK.E
	Laminin beta-3 chain	1	R.QTACTPGDCPGELCPQDN#GTACGSHCR.G
	Fc receptor, IgG, low affinity IIb	1	R.YHHYSSN#FSIPK.A
	Myelin P0 protein	1	K.DGSIVIHNLDYSDN#GTFTCDVK.N
	Ceruloplasmin	1	K.EYEGAVYPDN#TTDFQR.A
	Alpha-1-antitrypsin 1-6	1	K.GDTHTQILEGLQFN#LTQTSEADIHK.S
	Paired amphipathic helix protein Sin3a		P.DAN#SSVLLSKTTAEK.V
IPI00117957		1	R.ITDIEN#GTFANIPR.V
	mannosidase, beta A, lysosomal		V.AEILFNN#VTIGK.T
	Alpha-1-acid glycoprotein 1	1	R.ESQTIGDQCVYN#STHLGFQR.E
	Alpha-1-acid glycoprotein 1	1	R.QAIQTMQSEFFYLTTNLIN#DTIELR.E
	Alpha-1-acid glycoprotein 1	1	R.EN#GTFSKYEGGVETFAHLIVLR.K
	Receptor-type tyrosine-protein phosphatase N2		K.VSANIQN#MTTADVIK.A
	Glutamate [NMDA] receptor subunit zeta 1	1	K.VICTGPN#DTSPGSPR.H
	Complement component C8 gamma chain homolog	1	R.EAN#LTEDQILFFPK.Y
	Hypothetical Lipolytic enzyme, G-D-S-L containing		R.KGPGMENPVAVTIFFGAN#DSSLK.D
	protein	0.91	
	Leukemia inhibitory factor receptor	1	K.VVLAGSN#MTICCMSPTK.V
IPI00119299	Leukemia inhibitory factor receptor	1	R.IEGLTN#ETYR.L
	Leukemia inhibitory factor receptor	1	R.LGVQMHPGQEIHN#FTLTGR.N
	Carboxypeptidase N, polypeptide 2 homolog	1	R.LQDLEITGSPVSN#LSAHIFSN#LSSLEK.L
	Insulin receptor substrate 1	0.93	K.LLPCTGDYMN#MSPVGDSN#TS.S
			R.VPNNALEGLEN#LT.A
IPI00120187	FIDFOIIIOdulin	0.97	K.VFININALEULEIN#LI.A

IPI	Protein name	Р	Identified sequences
IPI00120769	Solute carrier family 29 (nucleoside transporters),	1	R.LDVSQN#VSSDTDQSCESTK.A
	member 1		
IPI00120848			I.SSLTDDTFCKAN#DTR.Y
	Versican core protein	1	R.FEN#QTCFPLPDSR.F
	Procollagen, type V, alpha 2	1	K.EASQN#LTYICR.N
IPI00121312		1	K.IDLTDFEKN#SSFA.Q
	F11r protein Retinoblastoma-associated protein	1	R.AFMN#SSFTIDPK.S K.QLEN#DTRIIEVLCKEHECNIDEVKN.V
	Sodium/potassium-transporting ATPase beta-1 chain	0.97	K.LDWLGN#CSGLNDDSYGYR.E
	High-affinity cationic amino acid transporter-1		K.FLAKINN#RTKTPVIATVTSGAIAAVM.A
IPI00121034 IPI00122293		1	R.VPVIPPRIHYLYLQNNFITELPLESFQN#ATGLR.W
	Neutrophil elastase homolog		R.LGTNRPSPSVLQELN#VT.V
	P2X4c receptor subunit	1	K.TSICDSDAN#CTLGSSDTHSSGIGTGR.C
IPI00122438	-	1	K.AWGTPCELCPSVN#TSEYK.I
IPI00122438		1	V.DTDECSVGNPCGN#GTCK.N
PI00122438		1	V.N#VTDYCQLVR.Y
PI00122438		1	R.NYYADN#QTCDGELLFN#MTK.K
IPI00122438		1	R.N#CTDIDECR.I
IPI00122456		1	R.MIEN#GSLSFLPTLR.E
IPI00123196		1	K.LGLSFNSITVMEN#GSLANVPHLR.E
IPI00123196		1	K.YIQVVYLHNNN#ISAVGQNDFCR.A
	Murinoglobulin-1	1	R.NYEVQLFHVN#ATVTEEGTGLEFSR.S
	Murinoglobulin-1	1	R.N#ASFVYTK.A
	Amiloride-sensitive sodium channel beta-subunit	1	K.GEPYSPCTMN#GSDVAIK.N
	Cd97 protein	1	R.DFNPATVN#YTIQK.L
	Neuropilin-1	1	K.RGPECSQN#YTAPTGVIK.S
	Macrophage scavenger receptor types I and II	1	R.VLNN#ITNDLR.L
	Osteoclast-like cell cDNA, clone:I420031M06	1	K.SDTPCDDFTRCPTN#NTCCK.L
11100121010	product:granulin	1	
IPI00124830	Leukocyte surface antigen CD47	1	I.EFTSCN#ETVVIPCIVR.N
	Laminin alpha-3 chain	1	K.IESINQQLLPLGN#ISDNVDR.I
	Laminin alpha-3 chain		K.TTFNLN#TTEVEPCRR.R
	Acid ceramidase	1	R.SVLEN#TTSYEEAK.N
	Eosinophil cationic protein 1		R.VHITVCN#ITSR.A
	Complement C1q subcomponent, A chain	1	K.VLTNQESPYQN#HTGR.F
	Peroxisomal 2,4-dienoyl-CoA reductase		F.RDHGGVIVN#ITATLSMR.G
	Ectonucleoside triphosphate diphosphohydrolase 5	1	R.GYLTSFEMFN#STFK.L
	Hypothetical protein	1	N.YQN#NTEVIQGIR.T
	Hypothetical protein	1	R.GLTFLKN#VSSTCAASPSTDILTFTIPPSFADIFLSK.S
	Plasma glutamate carboxypeptidase	1	K.EVMNLLQPLN#VTK.V
	Macrophage mannose receptor 1	1	R.TSYCN#ESFYFLCK.K
	Alpha-2-macroglobulin	1	K.N#ITSVVSPLGYLSIFTTDEHGLAN#ISIDTSN#FTAPFLR.
	Alpha-2-macroglobulin	1	R.IN#VSYTGERPSSNMVIVDVK.M
	Alpha-2-macroglobulin	1	Y.LN#ETQQLTEAIK.S
	Alpha-2-macroglobulin	1	K.VN#LSFPSAQSLPASDTHLK.V
	Mast cell carboxypeptidase A	1	R.NQN#STCIGTDLNR.N
	Vascular cell adhesion protein 1	1	K.ETTIWVSPSPILEEGSPVN#LTCSSDGIPAPK.I
	Myeloid bactenecin	1	K.DCDFLEDGEERN#CTGK.F
	AMBP protein	1	K.EDSCQLN#YSEGPCLGMQER.Y
	Transthyretin	1	K.TLGISPFHEFADVVFTAN#DSGHR.H
	PREDICTED: hypothetical protein LOC66967	1	K.LLPAFN#TTSGLPYPR.I
	Alpha-1-acid glycoprotein 2	1	R.EYHTIDDHCVYN#STHLGIQR.E
	Alpha-1-acid glycoprotein 2	1	D.PITN#ETLSWLSDK.W
	Androgen binding protein alpha	1	R.KVDLFLN#GTTEEY.V
	Alpha-2-HS-glycoprotein	1	R.RPFGVVYEMEVDTLETTCHALDPTPLAN#CSVR.Q
	r 0-7	-	

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IPI	Protein name	Р	Identified sequences
IPI00128484	Hemopexin	1	R.VAEVEN#GTKPD.S
IPI00128484	*	1	R.SWSTVGN#CTAALR.W
IPI00128484	-	1	K.SLGPNTCSSN#GSSLYFIHGPNLYCYSSIDK.L
IPI00128484	-	1	M.DHN#GTMLFFK.G
	Golgi phosphoprotein 2	1	K.AVLVNN#ITTGEK.L
	Vacuolar ATP synthase subunit S1	1	A.IHPPVSYN#DTAPR.I
	Tyrosine-protein phosphatase non-receptor type substrate 1	1	R.GIAN#LSNFIR.V
IPI00129243	Gamma-glutamyl hydrolase	1	K.LPLN#FTEGAR.K
	Gamma-glutamyl hydrolase	0.99	L.ALEN#LTANFHK.W
IPI00129250	Leucine-rich alpha-2-glycoprotein	1	L.SVEFSN#LTQLPAAALQGCPGLR.E
IPI00129250	Leucine-rich alpha-2-glycoprotein	1	K.MFSQN#DTR.C
IPI00129359	zinc finger protein 68	0.97	K.ELAGIGNTCN#VSTNH.I
IPI00129965	PREDICTED: similar to alpha-1-B glycoprotein	1	K.LLFVGPQHAGN#YSCR.Y
	PREDICTED: similar to alpha-1-B glycoprotein	0.99	R.VYQPGN#YSCSYQTHGECTSSTPSR.I
IPI00129968		1	K.DDEPLETTGDFN#TTK.M
	Complement factor H	1	K.DNSCVDPPHVPN#ATIVTR.T
	Complement factor H	1	K.LTEFTHN#STMDYK.C
	Complement factor H	1	R.TKCIN#GTINYPTCV
	Dipeptidyl-peptidase I	1	R.ILTN#NSQTPILSPQEVVSCSPYAQGCDGGFPYLIAGK.Y
	KH domain RNA binding protein QKI-5A		R.KDMYN#DTLN#GSTEK.R
IPI00130627	·		Y.DDIANSEEN#PTPGVVINRPN#GTDVYK.G
	Glutamate carboxypeptidase II	1	K.VPYNVGPGFAGN#FSTQK.V
IPI00130654		1	P.TKPODVDHFN#ATOK.F
IPI00130654 IPI00130654			L.ADLVLGELCGVNTN#R.T
		1	
	Tripeptidyl-peptidase I		K.DVGSGTTN#NSQACAQFLEQYFHNSDLTEFMR.L
	Tripeptidyl-peptidase I	1	K.SSSHLPPSSYFN#ASGR.A
	Type VI collagen alpha 3 subunit	1	R.GPPGVN#GTQGFQGCPGQR.G
	Type VI collagen alpha 3 subunit	1	R.ALN#GSALYTGSSLDFVR.N
	Type VI collagen alpha 3 subunit	1	R.QLINALQIN#NTAVGHALVLPAR.R
	9 kDa protein		K.GKAN#ASEDANNPAENGDAK.T
	Keratin intermediate filament 16a	1	R.KTEELNKEVASNSDLIQSN#R.S
	Keratin, type II cytoskeletal 6B	1	R.VPGLN#RSGFSSVSVCR.S
	CD209 antigen-like protein B	1	R.IPIFQGQN#ESIQEK.I
	Serine protease inhibitor A3K	1	K.NLINDYVSN#QTQGMIK.E
	Serine protease inhibitor A3K	1	K.YTGN#ASALLILPDQGR.M
IPI00131951	*	0.91	L.SLGAQN#STLEEIR.E
	NAD(P)(+)-arginine ADP-ribosyltransferase	1	R.LGN#FTLAYSAKPETADNQR.V
IPI00133035	NAD(P)(+)-arginine ADP-ribosyltransferase	1	K.GTSNDLVLQSIN#STCSYYECAFLGGLK.T
IPI00133172	Serpin B11	1	K.N#SSECSQVGVMHPDFR.A
IPI00133257	Hematopoietic progenitor cell antigen CD34	1	M.VLAN#STELPSK.L
IPI00133751	Microfibril-associated glycoprotein 4	1	R.VDLEDFEN#NTAYAK.Y
IPI00133751	Microfibril-associated glycoprotein 4	1	R.FN#GSVSFFR.G
IPI00134191	Solute carrier family 2, facilitated glucose transporter member 3	1	K.DFLN#YTLEER.L
IPI00134483	Lectin lambda	1	R.PGACTN#ITMGVVCK.L
IPI00134483	Lectin lambda	1	R.VTPVCN#ASLPAQR.W
	Zinc finger autosomal protein	0.98	V.ELLDPN#NSICVPREK.M
	Type VII collagen	1	K.LQILN#ASSDVLR.V
	C4b-binding protein	1	R.LACLN#GTVLR.G
	C4b-binding protein	1	R.LVGSPFIGCTVVN#K.T
	Antithrombin-III	1	K.LGACN#DTLK.Q
	Piccolo protein		Y.RRQISAVQPSIIN#LSAASSLGTPVTMDSK.T
	Immunoglobulin J chain	1	R.EN#ISDPTSPLR.R
	Lysosomal protective protein	1	R.LDPPCTN#TTAPSNYLNNPYVR.K
	Zinc-alpha-2-glycoprotein	1	K.DTTGSHTFQGMFGCEITNN#R.S
11 10013/30/	Zine-aipha-2-grycoprotein	1	K.D.1.05HITQUMEOCETINN#R.5

IPI	Protein name	Р	Identified sequences
	Liver carboxylesterase N	1	R.FHSELN#ISESMIPAVIEK.Y
IPI00139788	Serotransferrin	1	K.N#STLCDLCIGPLK.C
PI00153187	Sulfatase modifying factor 1	1	K.FVN#STGYLTEAEK.F
	Angiotensin-converting enzyme 2	0.99	Y.FFVTSPQN#VSDVIPR.S
	Protein Z-dependent protease inhibitor	1	R.ASQQLSN#ETSSFGFNLLR.K
	Hypothetical protein	0.9	C.QFGVGTFANVFLFVYN#FSPISTGSK.Q
	Procollagen, type VI, alpha 2	1	R.GTFTDCALAN#MTQQIR.Q
	Procollagen, type VI, alpha 2	1	I.GYTN#FTLEK.N
	Procollagen, type VI, alpha 2	1	R.MALLQYGSQNQQQVAFPLTYN#VTTIHEALER.A
	Procollagen, type VI, alpha 2 Procollagen, type VI, alpha 2	1	R.N#MTLFSDLVAEK.F
	Hypothetical protein LOC435366		
			R.HERN#QSAEKPSEYTQHGKAFALHAHSHAQ.R
	Choline transporter-like protein 2	1	K.TCNPETFPLRN#ESLQCPTAR.C
	Hypothetical Phospholipase D/Transphosphatidylase	1	K.VFIVPVGN#HSNIPFSR.V
	Glucosamine (N-acetyl)-6-sulfatase	1	K.YYN#YTLSINGK.A
	Synaptic vesicle glycoprotein 2 b		K.KVLSMSLAIN#ASFASLSSFVQGY.G
	Hypothetical Zinc finger, C2H2 type containing protein	0.96	D.WMPNN#HSVILIDDFESPQK.L
	Laminin alpha-4 chain	1	R.HVTDMN#STIHLLR.T
PI00223987	Insulin-regulated membrane aminopeptidase IRAP homolog	1	R.MAFDLIDYLKN#ETHTAPI.T
PI00224456	Sarcalumenin	1	A.PLIN#VTEPPR.V
	Sarcalumenin	1	K.TN#VSKFDLPNR.E
	Calsequestrin 2	1	K.IDLFKPQIGVVN#VTDADSI.W
	Hypothetical protein	1	R.AYIQDFQEFSKN#ISIMLGR.C
	Target of Nesh-SH3 variant 1		K.VHIN#TTSDSILLK.F
	Hypothetical von Willebrand factor type A domain	1	R.DLSVFAPN#MTEIIK.D
	containing protein	1	K.DL5 VFAFIN#IMTELIK.D
PI00226310	Hypothetical von Willebrand factor type A domain containing protein	1	K.LGN#FSELATHN#QTFLK.K
PI00226310	Hypothetical von Willebrand factor type A domain containing protein	0.99	L.LDMAIN#GSQEDLDHLK.A
PI00226790	GPI transamidase component PIG-T	0.92	L.GLAN#DTDDYFLR.Y
PI00226932	Quinoprotein alcohol dehydrogenase structure containing protein	1	R.FINYN#QTVSR.M
PI00227834	Inter-alpha trypsin inhibitor, heavy chain 2	1	K.GAFISN#FTMTVNGMTFTSSIK.E
	Hepatocyte growth factor activator	1	R.FCNIVPTEHCFLGN#GTEYR.G
PI00229117		1	Y.ILTYQFPN#GTVK.E
	Excitatory amino acid transporter 2	1	K.VLVAPPSEEAN#TTK.A
	PREDICTED: similar to type V P-type ATPase isoform 3		
	Huntington disease gene homolog		K.VCDPNSDVCN#TTR.S P. GVSLI PSITDVTMENNI#LSP V
			R.GYSLLPSITDVTMENN#LSR.V
	Murinoglobulin-2	1	K.ELIFYYLVMAQGSIIQTGN#HTHQVEPGEAPVK.C
	Proline 4-hydroxylase, alpha 1	1	K.DMSDGFISN#LTIQR.Q
PI00279010		1	F.VFLN#SSSTVVN#CSAR.G
	RIKEN cDNA A930025J12	1	R.LFQN#CSELYK.A
	Fibrinogen beta chain	1	K.GTAGNALMDGASQLVGEN#R.T
PI00281188	140 kDa protein	0.99	K.VLEPPHIN#GSEGPGEV.S
PI00281344	Hypothetical Glycosyl transferase, family 8 containing protein	0.93	R.TGVNSGVMLMN#MTR.M
	Ig gamma-1 chain C region, membrane-bound form	1	R.EEQFN#STFR.S
	Olfactomedin-like protein 3	1	K.IYVLDGTQN#DTAFVFPR.L
	E130014G12 product:Kaiso protein	1	K.EDLPSN#NT.A
	Serum amyloid P-component	1	K.LIPHLEKPLQN#FTLCFR.T
PI00309230	Beta-glucuronidase	0.98	R.ITIAIN#NTLTPH.T
PI00309999	Laminin alpha-2 chain	1	R.LEQMTMNIN#LTGPLPAPYK.I
PI00309999	Laminin alpha-2 chain	1	K.LN#ETLGNQDK.T
	-	1	R.ICNQN#SSNPYQR.H
PI00309999	Laminin alpha-2 chain	1	

IPI	Protein name	Р	Identified sequences
IPI00309999	Laminin alpha-2 chain	1	K.VCN#CSTVGSLASQCNVNTGQCSCHPK.F
	Laminin alpha-2 chain	1	K.ILYGLEN#TTQELK.H
IPI00309999	Laminin alpha-2 chain	1	K.YIGGGVCIN#CTHNTA.G
IPI00309999	Laminin alpha-2 chain	1	Y.VGGLPIN#YTTR.R
	Laminin alpha-2 chain	1	L.NLASNALITTN#ATCGEK.G
	Carboxypeptidase B2	1	K.EVHFFVN#ASDVDSVK.A
IPI00311808	Transmembrane glycoprotein NMB	1	R.DLPIVFDVLIHDPSHFLN#DSAISYK.W
IPI00313900		1	K.LHINYNN#LTESVGPLPK.S
IPI00313900	Lumican	1	R.LSHNELADSGVPGNSFN#ISSLLELDLSYNK.L
IPI00313900	Lumican	1	K.AFEN#VTDLQWLILDHNLLENSK.I
IPI00313900		1	K.LGSFDGLVN#LTFIYLQHNQLK.E
	keratin complex 2, basic, gene 1	1	R.MSGECTPN#VSVSVSTSHTSMSGSSSR.G
	T-cell immunomodulatory protein	1	V.PCNN#ASCEEVHR.M
	Adipocyte-derived leucine aminopeptidase	1	L.N#SSHPVSTPVENPAQIR.E
PI00319814	Suprabasal-specific protein suprabasin	1	K.EANQLLN#GSHQGQGGYGGQHGGAATT.T
IPI00320204	RIKEN cDNA 2210023G05	1	R.DGTSQPAICPQN#VTMNMEGLK.E
IPI00320675	Complement factor I	1	R.WGEVDLIGN#CSQFYPDR.Y
	Complement factor I	1	N.FN#VSLIYGR.T
IPI00321190	Sulfated glycoprotein 1	1	K.TN#SSFIQGFVDHVKEDCDR.L
IPI00321190	Sulfated glycoprotein 1	1	K.DN#ATQEEILHYLEK.T
IPI00322304	Histidine-rich glycoprotein HRG	1	R.LPPLNIGEVLTLPEANFPSFSLPNCN#R.S
IPI00322463	Beta-2-glycoprotein I	1	K.DYRPSAGN#NSLYQDTVVFK.C
IPI00322463	Beta-2-glycoprotein I	1	K.N#ISFACNPGFFLN#GTSSSK.C
IPI00322575	ATP-binding cassette transporter sub-family A member 8a	0.95	K.NTQNILVQN#LSGGQKRK.L
IPI00330747	5730439E10Rik protein	0.94	R.YLMGN#NSSEDSFLTANTVQPLAETGLQLSK.R
IPI00331214	Platelet glycoprotein IV	1	R.QFWIFDVQNPDDVAKN#SSK.I
IPI00331214	Platelet glycoprotein IV	1	K.DPFLSLVPYPISTTVGVFYPYN#DTVDGVYK.V
IPI00331214	Platelet glycoprotein IV	1	K.VISNN#CTSYGVLDIGK.C
IPI00331214	Platelet glycoprotein IV	1	K.RPYIVPILWLN#ETGTIGDEK.A
IPI00331214	Platelet glycoprotein IV	1	K.EN#ITQDPEDH.T
IPI00331214	Platelet glycoprotein IV	1	R.N#LSYWPSYCDMIN#GTDAASFPPFVEK.S
IPI00331259	Desmoglein-1 gamma	1	K.LN#ATDADEPNNLNSMIAFK.I
IPI00331617	Hypothetical olfactomedin-like domain containing protein	1	R.VDKLEEEVSKN#LTK.E
IPI00338565	Mutant fibrillin-1	1	R.VLPFN#VTDYCQLVR.Y
IPI00338785	CDNA, clone:M5C1012G13 product:laminin B1 subunit 1	1	K.MEMPSTPQQLQN#LTEDIR.E
IPI00338785	CDNA, clone:M5C1012G13 product:laminin B1 subunit 1	1	K.QADEDIQGTQNLLTSIESETAASEETLTN#ASQR.I
IPI00338785	CDNA, clone:M5C1012G13 product:laminin B1 subunit 1	1	L.ATGN#VSGGVCDNCQHNTMGR.N
IPI00338785	CDNA, clone:M5C1012G13 product:laminin B1 subunit 1	1	R.VN#ASTTDPN#STVEQSALTR.D
IPI00338785	CDNA, clone:M5C1012G13 product:laminin B1 subunit 1	1	K.LTDTASQSN#STAGELGALQAEAESLDK.T
	Collagen alpha 1(VI) chain	1	R.AALQFLQN#YTVL.A
	Collagen alpha 1(VI) chain	1	L.DDGFLKN#ITAQICIDKK.C
IPI00340463	PREDICTED: similar to hypothetical protein A030003A19	1	K.LLNDYVSN#QTQGMIK.E
	Spink5 protein	0.99	E.TNKNSASRSN#GTGSATGKDVCDQFR.S
	Spink5 protein		K.GNQDPCMKFQAQMKN#GTLTCPK.G
	Weakly similar to Zinnc finger protein GLI4	0.98	R.FRN#SSNLARHR.R
PI00350715	PREDICTED: similar to protocadherin 9	1	R.IDPVTGN#ITLEEKPAPTDVGLHR.L
PI00355606	PREDICTED: expressed sequence AL022779	0.96	M.QN#NSVFGDLK.S
IPI00378430	Ortholog of human Ras association	0.94	R.QETNMAN#FSYR.F
	Weakly similar to Tiarin	1	K.IN#LTTNVVDVNRPLPL.A
		1	M.IVNN#HTSLDVER.A
IPI00403586	Hypothetical Lipolytic enzymes	1	WI.IVINN#TISLDVEK.A

IPI	Protein name	Р	Identified sequences
IPI00406434	Mini-agrin	1	K.NELMLN#SSLMR.I
IPI00407222	PREDICTED: similar to human KIAA1815 protein	0.99	H.IPEIN#DTIR.A
	PREDICTED: similar to solute carrier family 4 member 11	0.94	R.EDSLGDEVFDTVN#SSIVSGESIR.F
IPI00409148	Haptoglobin	1	K.NLFLN#HSETASAK.D
IPI00409148		1	K.N#LTSPVGVQPILNEHTFCAGLTK.Y
IPI00409148	Haptoglobin	1	K.CVVHYEN#STVPEK.K
IPI00409148	Haptoglobin	1	K.VVLHPN#HSVVDIGLIK.L
IPI00410951	Thyroxine-binding globulin homolog	1	K.VTTCHLPQQN#ATLYK.M
IPI00420489	Von Willebrand factor	1	V.LEGSDEVGEANFN#K.S
IPI00420955	Sortilin 1	1	K.DITNLIN#NTFIR.T
IPI00453607	Killer cell inhibitory receptor-like protein p91A	1	R.LSVLPSPVVTAGGN#MTLH.C
	Sodium/glucose cotransporter 1	1	K.VSNGN#FTAK.E
	Kidney predominant protein	1	S.ADFQGRPVDDPTGAFAN#GSLTFK.V
IPI00460063	Prenylcysteine oxidase	1	K.GELN#STLFSSRPK.D
IPI00461281	NudC domain containing protein 2	1	K.ENPGFDFSGAEISGN#YTK.G
IPI00462999	Ahi-1 isoform III	0.92	D.EFVNTEN#NSSR.K
IPI00463311	PREDICTED: similar to RIKEN cDNA E330026B02	0.99	R.DLGMFAPN#MTR.I
IPI00467180	Translocon-associated protein beta subunit	1	R.IAPASN#VSHTVVLRPLK.A
IPI00467944	61 kDa protein	1	K.VVN#VSELYGTPCTK.R
IPI00468097	340 kDa protein	1	R.NLQVYN#ATSNSLTVK.W
IPI00469000	Zinc transporter SLC39A6	0.98	R.NTNDNIQECFN#TTK.L
	GUGU alpha	1	R.VLYLPAYN#CTLRPVSK.R
IPI00469387	GUGU alpha	1	R.SPPAPPLPQRPLSPLHPLGCN#DSEVLAVAGFALQNINR.D
IPI00469542	Histidine-rich calcium-binding protein	1	R.EVGEEN#VSEEVFR.G
IPI00469839	19 kDa protein	0.91	K.TRTIDVVYN#ASNNELVCTK.T
IPI00471081	RIKEN cDNA 1100001H23	1	K.NGDAYGYYN#DSIK.T
IPI00471273	Apoptosis-related protein 3	1	A.LPEICTLCPGGMHN#LSR.V
	PREDICTED: laminin, alpha 3	0.98	R.FN#ISTPAFQGCMK.N
IPI00473830	Biliary glycoprotein	1	R.MTLSQN#NSILR.I
IPI00475154	Dolichyl-diphosphooligosaccharide-protein glycosyltransferase 63 kDa	1	Q.VLSGCEISVSN#ETK.E
IPI00475157	Serpina1b protein	1	R.ELVHQSN#TSNIFFSPVSIATAFAMLSLGSK.G
IPI00475157	Serpinalb protein	1	N.ASAVFLLPEDGK.M
IPI00551354	PREDICTED: ring finger and KH domain containing 3	0.91	R.N#GSGGGGGGGGGGGGGGGGGGGGGGGGGGAA
IPI00553278	H-2D cell surface glycoprotein	1	R.NLLGYYN#QSAGGSHTLQQM.S
IPI00554833	Eosinophil-associated ribonuclease 12	1	V.GVCGN#PSGLCSDN#ISQNCHN#SSSR.V
IPI00606550	Ig gamma-2B chain C region, membrane-bound form	1	R.EDYN#STIR.V
IPI00607976	Serine (or cysteine) proteinase inhibitor, clade A, member 3A	1	K.FN#LTETPEADIH.Q
IPI00621319	43 kDa protein	0.92	K.RLFLLDLLN#ATGK.D
IPI00624663	*	0.99	K.ACVSLNHVN#ETVM.L
IPI00624761	44 kDa protein	1	R.PVDDPTGAFAN#GSLTFK.V
	38 kDa protein	0.94	P.PSSTDLLWSILN#ASALALLYKTQRDN#ASESK.D
	MKIAA4087 protein		R.CNIN#GSFSEICHTR.T
	Adult male thymus cDNA, clone:5830446P09 product: CD72 antigen	0.96	V.GSEQPTATWSSVN#SSALRQIPR.C
IPI00649281	52 kDa protein	0.98	R.YHYN#GTLLDGTAFDNSYSR.N
	Myosin light chain, regulatory B	0.91	K.N#PTDAYLDAMMNEAPAPIN#FTMFL.T
	Hypothetical protein CEACAM1/2sec	1	R.FHVHQPVTQPFLQVTN#TTVK.E

P: peptide probability N#: N-linked glycosylation site

Table 2 Glycoproteins upregulated in skin tumors

IPI	Protein name	Protein location	Ca	Ра	Nr	Total	Ca/Nr	Pa/Nr	Ca/Pa
IPI00119063	AM2 receptor	Transmembrane	17	0	0	17	100.0	0.0	100.0
IPI00381122	Weakly similar to Tiarin	Cell Surface	11	0	0	11	100.0	0.0	100.0
IPI00308971	Cation-independent mannose-6-phosphate receptor	Transmembrane	8	0	0	8	100.0	0.0	100.0
IPI00124265	Latent transforming growth factor beta binding protein 4	Secreted	7	0	0	7	100.0	0.0	100.0
IPI00129304	Collectin sub-family member 12	Transmembrane	7	0	0	7	100.0	0.0	100.0
IPI00129968	Embigin	Transmembrane	7	0	0	7	100.0	0.0	100.0
IPI00153959	Stabilin-1	Transmembrane	5	0	0	5	100.0	0.0	100.0
IPI00316575	Cathepsin K	Cell Surface	4	0	0	4	100.0	0.0	100.0
IPI00321190	Sulfated glycoprotein 1	Secreted	4	0	0	4	100.0	0.0	100.0
IPI00475154	Dolichyl-diphosphooligosaccharide-protein glycosyltransferase 63 kDa	Transmembrane	4	0	0	4	100.0	0.0	100.0
IPI00308785	Prostaglandin G/H synthase 2	Secreted	3	0	0	3	100.0	0.0	100.0
IPI00122737	222 kDa protein	Intracellular	3	0	0	3	100.0	0.0	100.0
IPI00406459	Arylsulfatase B	Secreted	3	0	0	3	100.0	0.0	100.0
IPI00409393	Latent transforming growth factor beta binding protein, isoform 1L	Secreted	3	0	0	3	100.0	0.0	100.0
IPI00119809	Mama protein	Secreted	3	0	0	3	100.0	0.0	100.0
IPI00111960	Lysosomal alpha-glucosidase	Transmembrane	3	0	0	3	100.0	0.0	100.0
IPI00118011	Mannosidase, beta A, lysosomal	Secreted	3	0	0	3	100.0	0.0	100.0
IPI00121120	Procollagen, type V, alpha 2	Secreted	3	0	0	3	100.0	0.0	100.0
IPI00129158	Tyrosine-protein phosphatase non-receptor type substrate 1	Transmembrane	3	0	0	3	100.0	0.0	100.0
IPI00120769	Solute carrier family 29 (nucleoside transporters), member 1	Transmembrane	5	0	1	6	5.0	0.0	100.0
IPI00415773	Integrin alpha-M	Transmembrane	34	3	1	38	34.0	3.0	11.3
IPI00338785	cDNA, clone:M5C1012G13 product:laminin B1 subunit 1	Intracellular	9	1	3	13	3.0	0.3	9.0
IPI00130627	Legumain	Secreted	17	2	0	19	100.0	100.0	8.5
IPI00113480	Myeloperoxidase	Secreted	8	1	0	9	100.0	100.0	8.0
IPI00113824	Basement membrane-specific heparan sulfate proteoglycan core protein	Cell Surface	15	2	5	22	3.0	0.4	7.5
IPI00124830		Transmembrane	7	1	1	9	7.0	1.0	7.0
IPI00320605	Integrin beta-2	Transmembrane	20	3	0	23	100.0	100.0	6.7
IPI00308990	Monocyte differentiation antigen CD14	Cell Surface	13	2	0	15	100.0	100.0	6.5
IPI00133082	CD177 antigen	Secreted	6	1	0	7	100.0	100.0	6.0
IPI00130486	FK506-binding protein 9	Cell Surface	5	1	0	6	100.0	100.0	5.0
IPI00308609	VESICULAR INTEGRAL-MEMBRANE PROTEIN VIP36	Transmembrane	5	1	0	6	100.0	100.0	5.0
IPI00403938	Tenascin-C	Cell Surface	133	29	0	162	100.0	100.0	4.6
IPI00462199	Basigin	Transmembrane	13	3	0	16	100.0	100.0	4.3
IPI00120245	Integrin alpha-V	Transmembrane	8	2	0	10	100.0	100.0	4.0
IPI00120245	Integrin alpha-V	Transmembrane	8	2	0	10	100.0	100.0	4.0
IPI00110852	Translocon-associated protein alpha, muscle specific isoform	Cell Surface	4	1	0	5	100.0	100.0	4.0
IPI00125266	Acid ceramidase	Secreted	4	1	0	5	100.0	100.0	4.0
IPI00121038	Versican core protein	Cell Surface	11	3	2	16	5.5	1.5	3.7
IPI00124283	Macrophage scavenger receptor types I and II	Transmembrane	7	2	0	9	100.0	100.0	3.5
IPI00132067	Fibulin-2	Secreted	31	9	0	40	100.0	100.0	3.4
IPI00223769	CD44 antigen	Transmembrane	10	3	0	13	100.0	100.0	3.3
IPI00127447	Lysosome membrane protein II	Transmembrane	32	10	0	42	100.0	100.0	3.2
IPI00322447	RA175	Transmembrane	6	2	0	8	100.0	100.0	3.0
IPI00154057	Protocadherin 1	Cell Surface	3	1	0	4	100.0	100.0	3.0
IPI00121312	MFIRE1	Secreted	3	1	0	4	100.0	100.0	3.0
IPI00124640 IPI00134483	Osteoclast-like cell cDNA, clone:I420031M06 product:granulin Lectin lambda	Secreted Cell Surface	3 3	1 1	0 1	4 5	100.0 3.0	100.0 1.0	3.0 3.0

IPI	Protein name	Protein location	Ca	Pa	Nr	Total	Ca/Nr	Pa/Nr	Ca/Pa
IPI00272381	Proline 4-hydroxylase, alpha 1	Secreted	17	6	0	23	100.0	100.0	2.8
IPI00122493	FK506-binding protein 10	Secreted	7	3	0	10	100.0	100.0	2.3
IPI00123831	SDR1 protein	Transmembrane	11	5	0	16	100.0	100.0	2.2
IPI00224728	Cd63 antigen	Transmembrane	8	4	0	12	100.0	100.0	2.0
IPI00128689	Collagen alpha 1(V) chain	Secreted	6	3	0	9	100.0	100.0	2.0
IPI00125877	Hypothetical protein	Transmembrane	6	3	0	9	100.0	100.0	2.0
IPI00130015	Dipeptidyl-peptidase I	Secreted	4	2	0	6	100.0	100.0	2.0
IPI00318012	T-cell immunomodulatory protein	Transmembrane	4	2	0	6	100.0	100.0	2.0
IPI00123678	Cadherin-22	Transmembrane	2	1	0	3	100.0	100.0	2.0
IPI00126316	Mast cell carboxypeptidase A	Secreted	2	1	0	3	100.0	100.0	2.0
IPI00130661	Tripeptidyl-peptidase I	Cell Surface	2	1	0	3	100.0	100.0	2.0
IPI00131366	Keratin, type II cytoskeletal 6B	Transmembrane	2	1	0	3	100.0	100.0	2.0
IPI00221418	hypothetical Phospholipase D/Transphosphatidylase	Transmembrane	2	1	0	3	100.0	100.0	2.0
IPI00279051	RIKEN cDNA A930025J12	Transmembrane	2	1	0	3	100.0	100.0	2.0
IPI00554833	Eosinophil-associated ribonuclease 12	Secreted	2	1	0	3	100.0	100.0	2.0
IPI00127280	Myeloid bactenecin	Secreted	43	22	0	65	100.0	100.0	2.0
IPI00118413	Thrombospondin 1	Secreted	20	11	0	31	100.0	100.0	1.8
IPI00127352	AMBP protein	Secreted	22	14	0	36	100.0	100.0	1.6
IPI00132600	Niemann-Pick C1 protein	Transmembrane	3	2	0	5	100.0	100.0	1.5
IPI00137177	Lysosomal protective protein	Secreted	3	2	0	5	100.0	100.0	1.5
IPI00132474	Integrin beta-1	Transmembrane	18	13	1	32	18.0	13.0	1.4
IPI00123342	Hypoxia up-regulated 1	Secreted	19	14	0	33	100.0	100.0	1.4
IPI00126090	Integrin alpha-3	Transmembrane	4	3	0	7	100.0	100.0	1.3
IPI00131881	ADAM 10	Cell Surface	4	3	0	7	100.0	100.0	1.3
IPI00406434	Mini-agrin	Secreted	4	3	0	7	100.0	100.0	1.3
IPI00410951	Thyroxine-binding globulin homolog	Secreted	4	3	0	7	100.0	100.0	1.3
IPI00125058	Laminin alpha-3 chain	Secreted	9	7	1	17	9.0	7.0	1.3
IPI00112326	Epithelial membrane protein 1	Transmembrane	6	5	0	11	100.0	100.0	1.2
IPI00128154	Cathepsin L	Secreted	23	20	0	43	100.0	100.0	1.2
IPI00121362	F11r protein	Transmembrane	9	9	0	18	100.0	100.0	1.0
IPI00108535	Carcinoembryonic antigen-related cell adhesion molecule 1	Cell Surface	7	7	0	14	100.0	100.0	1.0
IPI00407222	PREDICTED: similar to human KIAA1815 protein	Transmembrane	6	6	0	12	100.0	100.0	1.0
IPI00128989	Vacuolar ATP synthase subunit S1	Transmembrane	5	5	0	10	100.0	100.0	1.0
IPI00471081	RIKEN cDNA 1100001H23	Cell Surface	5	5	0	10	100.0	100.0	1.0
IPI00226932	Quinoprotein alcohol dehydrogenase structure containing protein	Secreted	4	4	0	8	100.0	100.0	1.0
IPI00127672	PREDICTED: hypothetical protein LOC66967	Secreted	2	2	0	4	100.0	100.0	1.0
IPI00346978	Spink5 protein	Secreted	2	2	0	4	100.0	100.0	1.0
IPI00469387	GUGU alpha	Secreted	23	23	3	49	7.7	7.7	1.0
IPI00134549	Lysosome-associated membrane glycoprotein 2	Transmembrane	8	9	0	17	100.0	100.0	0.9
IPI00121430	Collagen alpha 1(XII) chain	Secreted	11	14	0	25	100.0	100.0	0.8
IPI00122272	Extracellular matrix protein 1	Secreted	11	14	0	25	100.0	100.0	0.8
IPI00227969	Integrin alpha-6	Transmembrane	6	8	1	15	6.0	8.0	0.8
IPI00134652	Type VII collagen	Secreted	5	7	0	12	100.0	100.0	0.7
IPI00114256	Synaptophysin-like protein	Transmembrane	10	14	3	27	3.3	4.7	0.7
IPI00110810	Prostate stem cell antigen	Secreted	9	13	0	22	100.0	100.0	0.7
IPI00467180	Translocon-associated protein beta subunit	Transmembrane	15	22	0	37	100.0	100.0	0.7
IPI00133172	Serpin B11	Intracellular	2	3	0	5	100.0	100.0	0.7
IPI00111013	Cathepsin D	Secreted	19	30	0	49	100.0	100.0	0.6
IPI00117093	Laminin beta-3 chain	Cell Surface	3	6	0	9	100.0	100.0	0.5
IPI00130342	Lymphocyte antigen 6 complex locus G6C protein	Secreted	2	4	0	6	100.0	100.0	0.5
IPI00125293	Eosinophil cationic protein 1	Secreted	1	2	0	3	100.0	100.0	0.5
IPI00123293 IPI00320204	RIKEN cDNA 2210023G05	Secreted	1	2	0	3	100.0	100.0	0.5
IPI00320204 IPI00468097		Secreted	4	2 8	1		4.0	8.0	0.5
	340-kDa protein					13			
IPI00113853	Desmocollin-3	Transmembrane	2	6	0	8	100.0	100.0	0.3

Table 2 ((continued)
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IPI	Protein name	Protein location	Ca	Ра	Nr	Total	Ca/Nr	Pa/Nr	Ca/Pa
IPI00319814	Suprabasal-specific protein suprabasin	Secreted	3	10	0	13	100.0	100.0	0.3
IPI00115854	TROP2 protein	Transmembrane	1	4	0	5	100.0	100.0	0.3
IPI00127933	Androgen binding protein alpha	Secreted	1	4	0	5	100.0	100.0	0.3
IPI00130249	GPI-anchored metastasis-associated protein homolog	Secreted	10	60	0	70	100.0	100.0	0.2
IPI00111014	Elongation of very long chain fatty acids protein 4	Transmembrane	3	20	2	25	1.5	10.0	0.2
IPI00129243	Gamma-glutamyl hydrolase	Secreted	1	8	0	9	100.0	100.0	0.1
IPI00338790	Glandular kallikrein KLK13	Cell Surface	0	6	0	6	0.0	100.0	0.0
IPI00111115	Similar to METASTASIS-ASSOCIATED GPI-	Secreted	0	4	0	4	0.0	100.0	0.0
	ANCHORED PROTEIN								
IPI00473830	Biliary glycoprotein	Transmembrane	0	4	0	4	0.0	100.0	0.0
IPI00153548	Hypothetical protein	Transmembrane	0	3	0	3	0.0	100.0	0.0

Ca/Nr: Ratio of spectral count of carcinomas to normal tissue, Pa/Nr: Ratio of spectral count of papillomas to normal tissue, Ca/Pa: Ratio of spectral count of carcinomas to papillomas

Detected Tissue-Derived Proteins in Plasma

Since the plasma proteome is dominated by several highly abundant proteins, proteins released from specific tissues would normally be present at low abundance in plasma, and their detection might be obscured by the highly abundant plasma proteins. To detect tumor-specific proteins in plasma, we used isotopic labeling to detect the isotopic peaks that consisted of the tissue-derived proteins from both plasma and tissues.

The glycopeptides from four carcinomas were labeled with $d4^{13}C4$ -succinic anhydride. The glycopeptides from plasma of the four mice before and after cancer development were labeled with $d0^{13}C0$ and $d4^{13}C0$ -succinic anhydride, respectively. To monitor the labeling efficiency, we spiked the same amount of standard peptide from angiotensin (0.1 µg) in the glycopeptides isolated from carcinomas and plasma samples as labeling control. Then, all the labeled peptides were combined for MS analysis. The mixture was separated by 2D Nano-LC then analyzed by MALDI-TOF/TOF. Free angiotensin (ms 1296.68) was not observed after labeling. Instead, 100, 104, and 108 Da shifted from 1296.68 were observed in equal amounts in the mixed samples. This indicates the efficient and quantitative isotopic labeling using succinic anhydride.

The mixed glycopeptides from carcinomas and plasma samples contained both skin-cancer-related peptides and peptides from plasma. In order to detect glycopeptides associated with skin cancer in plasma, we focused our analysis on glycopeptides previously identified as cancerassociated glycoproteins from skin tumors in the mixture (Table 2) and avoided the analysis of plasma proteins. To achieve this goal, the peptide peaks that contained masses from glycopeptides specifically identified from carcinomas and their isotopic pairs from plasma were selected for MS/ MS analysis. Two types of paired patterns were observed. One was that the intensity of $d4^{13}C4$ -labeled peptides (with 8 mass unit shift for each amino group from peptides derived from cancer tissues) was much greater than $d4^{13}C0$ -labeled peptide (with 4 mass unit shift for each amino group from peptides derived from plasma of cancer-bearing mice), and intensity of $d0^{13}C0$ -labeled peptide (with 0 mass unit shift for each amino group from peptides derived from plasma of cancer-bearing mice), and intensity of $d0^{13}C0$ -labeled peptide (with 0 mass unit shift for each amino group from peptides derived from plasma before carcinogen induction) was lower than that of peptides from plasma of cancer-bearing mice. This pattern indicated that the peptide was from tumor-specific protein and detectable in cancer plasma at low intensity. The other pattern was that similar or lower intensity of peptides from cancer tissues than in plasma, and peptides with this pattern were derived from plasma proteins.

Tumor-associated glycopeptides could be detected in plasma. Tenascin-C was identified in carcinomas with 133 spectra, and it was also identified in benign papillomas with 29 spectra. However, none of these glycopeptides were identified in normal tissue (Table 2). In plasma, the labeled peptide peak of Tenascin-C from cancer was found with its paired peak from cancer plasma (Fig. 2A), which indicated that it was also detected in plasma after cancer development, but not in control plasma before the carcinogen treatment. Another skin tumor-specific glycoprotein, Arylsulfatase B, was also detected in plasma successfully in a similar way (Fig. 2b). These data indicated that extracellular proteins associated with tumor development were identifiable in plasma from tumor-bearing mice using glycopeptide capture, isotopic labeling, and mass spectrometry.

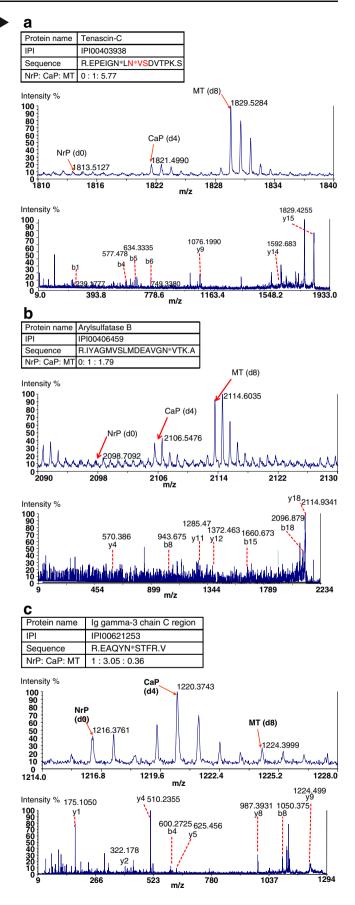
One of the advantages of using this tissue-targeted approach is that tumor-associated proteins can be identified in plasma even if they are present in very low abundance. The peptides from cancer tissue are likely to be at higher abundance compared to the same peptides in plasma. This allowed us to determine their masses and peptide sequences in the mixture

Fig. 2 Detection of tumor-specific proteins in plasma. **a** The detected paired peaks of succinic-anhydride-labeled Tenascin-C and MS/MS spectrum of Tenascin-C. **b** The paired peak of succinic-anhydride-labeled Arylsulfatase B and MS/MS of Arylsulfatase B. **c** The paired peak of succinic-anhydride-labeled Ig gamma-3 chain C region showed different peak pattern from Tenascin-C and MS/MS of Ig gamma-3 chain C region. *NrP* mouse plasma without carcinogen treatment, *CaP* mouse plasma from cancer-bearing mice after carcinogen treatment, *MT* mouse cancer tissues

using isotopic peaks from tumors. Using this information, tumor-derived peptides in plasma can be identified, while they are not identifiable by data-dependent MS/MS acquisition and database searches. Both Tenascin-C and Arylsulfatase B are low abundant proteins. They were not identified in plasma before cancer development, and their detection in plasma was associated with cancer development.

Proteins from plasma can also be detected in tissues and plasma as isotopic pairs due to vascularization of the tissue. If a glycopeptide detected in both cancer tissues and plasma was derived from plasma, the peptide peak showed similar or lower intensity in cancer tissues than that in plasma. An example of this was the identification and quantification of glycoprotein, Ig gamma-3 chain C region, in tissue and plasma. However, its paired peptide peaks were found in a different pattern from that observed with Tenascin-C (Fig. 2c). The intensities of $d0^{13}$ CO-and $d4^{13}$ CO-labeled peptides from plasma before and after tumor induction were much higher than that from $d4^{13}$ C4-labeled peptides from tumors. This indicated that this peptide was from a plasma-derived protein, and Ig gamma-3 could be detected from tissue due to the blood contamination in the tumor.

The methodology of targeted detection of tumor proteins using glycopeptide capture, isotopic labeling, and mass spectrometry is based on the analysis of N-linked glycopeptides to study extracellular proteins from tumors and plasma. It has been shown to increase the detectability of tumor proteins by focusing on the same subset of glycopeptides in both tumors and plasma [13]. The tumorassociated glycopeptides could be detected in plasma on account of the several advantages of our methodologies. First, the glycopeptide capture method dramatically reduces the sample complexity. Non-glycoproteins and non-glycopeptides from glycoproteins were removed from the pool of samples. For example, albumin, the most abundant serum protein, was automatically transparent to this method since it does not contain N-linked glycosylation. Second, the glycopeptide isolation method could be used to enrich extracellular proteins due to the fact that most extracellular proteins are glycosylated and likely to enter the bloodstream. Third, we used an isotopic labeling method to facilitate the detection of tumor proteins within complex plasma by identifying paired peptide peaks from tumor tissues and plasma. However, the method described



here is only for proteins that contain N-linked glycosylation. For proteins that do not contain N-linked glycosylation, this method will miss the detection of those proteins.

These results show that our strategy for detection of tumor-specific proteins in plasma is specific and sensitive for low abundant tumor-associated proteins. Different from the previous report of identification of prostate cancer-derived proteins in serum using xenograft-bearing mice [30], our study is more focused on tumor-associated extracellular proteins that are likely to be used in early detection.

Conclusions

In this study, we described a platform for quantitative detection of tumor-specific extracellular proteins derived from tumors and plasma. The fact that tumor-specific proteins were detectable in plasma from tumor-bearing mice indicates that cancer-specific markers could be detected in plasma using targeted approaches, and these proteins could be serum tumor marker candidates [7]. Once such candidate proteins are identified, the homologues of the proteins can be verified in human sera using the targeted approach. Enzyme-linked immunosorbent assays can be developed using a pair of antibodies. However, if antibodies against the candidate proteins are not available, mass-spectrometry-based methods can be applied to detect candidate proteins in plasma. One approach is referred to as MRM [17-19]. In another approach, called stable isotope standards and capture by anti-peptide antibodies, a specific peptide from sample and the synthetic heavy isotope-labeled peptide of the candidate protein are captured by peptide antibody. The mass spectrometer is then used to detect and quantify the specific peptide with known precursor mass and fragmentation ions [31].

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